

## RNA Confirmation Procedure

### **Step One: Design Primers**

- Using sequence from CMHD database and NeoL primer, design R primer from web resource such as Primer3
- Sequence from database should be blasted in NCBI/ensembl before primer is designed to make sure the sequence is good
- Using sequence of gene and new primer, determine expected length of PCR product

### **Step Two: RNA isolation/purification**

- RNA is purified from expanded ES cells using the Machery-Nagel RNA kit

### **Step Three: cDNA synthesis**

- Protocol follows

### **Step Four: PCR**

- Protocol follows

### **Step Five: Check PCR products**

- Run pcr products on 1.5% agarose gel

### **Step Six: Send for Sequencing**

- If the PCR band is of expected size, send PCR product for sequencing using the primer listed in section below.

### **Step Seven: Blast sequence results against CMHD and NCBI/ensembl databases.**

- When sequencing is returned, blast sequence against CMHD database to determine if the new sequence matches the old sequence
- Also, blast the new sequence through NCBI/ensembl to see if the correct gene is sequenced

# RNA Confirmation Protocol

## **Part One: cDNA Synthesis**

*(Invitrogen M-MLV RT protocol)*

1. Mix together:
  - 1 ul 3'CDS primer
  - 1 ul dNTP mix
  - 2 ul purified RNA
  - 8 ul H<sub>2</sub>O

The 3CDS primer sequence is AAG CAG TGG TAA CAA CGC AGA GTA CTT TTT TTT TTT TTT TTT TTT TTT TVN

2. Heat mixture to 65°C for 5 minutes (*PCR program 655min*). Chill on ice immediately afterwards.
3. Add:
  - 1 ul RNase OUT
  - 2 ul DTT
  - 4 ul 5x First-strand Buffer
4. Mix and incubate at 37°C for 2 minutes (*PCR program 3702min*).
5. Add in 1 ul of M-MLV RT buffer.
6. Incubate 37°C for 50 minutes (*PCR program RTLIA*).
7. Inactivate reaction by incubating at 70°C for 15 minutes.

## **Part Two: PCR**

*(Invitrogen M-MLV RT protocol)*

### Master Mix:

ddH <sub>2</sub> O	38.0ul
10x PCR Buffer	5.0ul
MgCl	1.5ul
dNTP mix	1.0ul
Left Primer	1.0ul
Right Primer	1.0ul
Taq	0.5ul
cDNA	2.0ul
<b>Total</b>	<b>50.0ul</b>

### Cycling Conditons:

94°C	2 minutes
94°C	30 seconds
62°C	30 seconds
72°C	2 minutes
repeat steps 2-5 for 39 cycles	
72°C	2 minutes
hold at 4°C	

**Program: RHOATR**

Left primer (F) is listed below. Right primer (R) is your gene specific primer that you design.

## Primers for Clone Confirmation

### pGTlox4/4pA

- Left: (bsdL) GTTATGTGTGGGAGGGCTAAGCAAT
- 69bp to splice site
- Sequencing: bsdL GTTATGTGTGGGAGGGCTAAGCAAT

### SD5

- Left: (NeoL) GCTATCAGGACATAGCGTTGGCTAC
- 193bp to splice site
- Sequencing: NeoL :

### pMS

- Left: (NeoL) GCTATCAGGACATAGCGTTGGCTAC
- 385bp to splice site
- Sequencing: NeoL

### NMDi

- Left: (NeoL) GCTATCAGGACATAGCGTTGGCTAC
- 192bp to splice site
- Sequencing: NeoL

### pUPA

- Left: (NeoL) GCTATCAGGACATAGCGTTGGCTAC
- 918bp to splice site
- Sequencing: (UPAtrap 7) ATTCCAGACAAGTTTGTGTTGGATATGCC
- 64bp to splice site